UC Davis Aquatic Health Program Laboratory



	Progress Report #	1							
	Reporting Period:	Nov. 1, 2021 to Jan. 31, 2022							
	Submittal Date:	February 10, 2022							
Contract No:	Department of Pesticide Regulation No. 21-C	Department of Pesticide Regulation No. 21-C0044							
Project Name:	Evaluation of Pyrethroid Toxicity Removal in Hyalella azteca	Evaluation of Pyrethroid Toxicity Removal in Agricultural Detention Basins using Hyalella azteca							
Contractor Name:	UC Davis Aquatic Health Program Laboratory								

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Project Director:	Dr. Swee Teh	Svee Til
	Printed Name	Signature

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Report Narrative

Introduction

Retention basins and other wetland-based management practices have been shown to be effective in removing pyrethroid pesticides from surface water. This water treatment practice is likely to be successful in removing pyrethroids from agricultural runoff, however, research on pyrethroid removal and mitigation of pyrethroid toxicity has been limited. The Department of Pesticide Regulation has contracted with Dr. Tom Young of UC Davis to determine the distribution and settling properties of particle suspensions of pyrethroids in field collected agricultural samples. This project will focus on quantifying pyrethroid toxicity removal in the process of sedimentation using pyrethroid sensitive testing organism *Hyalella azteca* (*H. azteca*).

Summary of Activities

The contract was approved on November 5, 2021. All parties held a project kick-off meeting on November 10, 2021, to discuss the logistics of the project and to work out sampling plans. It was decided that half of the

samples would be collected at this time, in part due to the delay in getting the contract in place, which put us outside the irrigation season. It was also decided to do a partial collection in order to optimize the protocols to be used for the project and to ensure that the DOC/TOC were at the proper ratio for testing. Samples were collected from Sal_Hartnel (Site 1) on November 29, 2021 and from Sal_Tembl (Site 2) on November 30, 2021. Toxicity tests were initiated on December 1, 2021.

Materials and Methods

Water Sample Collection

For *H. azteca* toxicity tests, 3L of whole water (WW) was collected from each site as sub-surface grabs via a pump. Three (3)-L of clarified supernatant (CS) was collected via a flow-through centrifuge from each site. These samples were collected in pre-cleaned 1L amber bottles. An additional 240L of WW were collected in 12 -20L cubtainers for future settling tests. All containers used for water collections were labeled with the site ID, collection date and time, initials of the sampler and then rinsed three times with ambient water prior to filling. All samples were placed on wet ice for transport to the UCD AHP and kept between 0-6°C³. Upon receipt, samples were stored in the dark in an environmental chamber maintained between 0-4°C until their use in a test.

Water Quality

Ammonia-nitrogen was measured at UCD AHPL within 24 hours of sample receipt using a HACH DR-3900 spectrophotometer and a HACH Am-Ver Low-Range Ammonia Reagent Set. Hardness and alkalinity were measured on all ambient samples (titrimetric methods) within 48-hours of sample receipt. These measurements were taken on the control and the 100% concentration of each ambient sample (whole water and clarified supernatant) only.

Toxicity Testing Methods – Acute Hyalella

UCD AHP toxicity testing methods are based on protocols developed by UCD AHP SOPs¹, SWAMP QAPrP ², and USEPA³. Acute toxicity testing for *H. azteca* followed protocols outlined in *Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms*³ and SWAMP Method Quality Objectives².

Before test initiation and water renewals, water samples were shaken thoroughly in their original sample containers for 60 seconds. Prior to test initiation and renewals, waters were warmed to test temperature (23 \pm 1°C using a water bath maintained at 25 \pm 2°C and aerated at a rate of 100 bubbles per minute until the DO concentration fell below saturation. Water quality measurements including pH, Specific Conductivity (SC), Dissolved Oxygen (DO) and temperature were recorded for all treatments at test initiation, water renewals, and termination.

H. azteca were obtained from Aquatic Research Organisms (Hampton, NH) and were acclimated to laboratory conditions for 24-h prior to test initiation. Acute 96-h toxicity tests consisted of five 250 mL replicate glass beakers with 100 mL of sample, a $1-in^2$ of nitex screen for artificial substrate, and 10 organisms each. Samples were tested in a dilution series consisting of five concentrations of either WW or CS, and a control. Reverse-Osmosis water amended with inorganic salts to USEPA moderately hard standards (ROEPAMHR) was used as the control and as dilution water. Eighty percent of the test solution was renewed at the 48-hr timepoint. Organisms were fed 1.5 mL of YCT (a mixture of yeast, trout chow, organic alfalfa and water) after test initiation and water renewal. Tests were conducted at $23 \pm 1^{\circ}$ C with a 16-hr light: 8-hr dark photoperiod under

fluorescent light. Mortality was scored daily; at this time dead organisms and detritus were removed from test chambers if present. A reference toxicant test with sodium chloride as the toxicant, was conducted currently with each set of toxicity tests.

Statistics

Each sample was characterized by descriptive statistics, including the mean response and variation among replicates. Toxicity is defined as a statistically significant reduction in test organism performance in an ambient sample compared to a laboratory control. Lethal effect concentrations were calculated using CETIS v. 1.8.7.2 (Tidepool Scientific Software, McKinleyville, CA, USA). NOEC and LOEC values were calculated using USEPA standard statistical protocols. LC50s were calculated using linear regression.

Results

Ambient samples were collected on November 29 and 30, 2021 and applied in toxicity tests with *Hyalella azteca* on December 1, 2021. Statistically significant reductions in survival were observed in the 100%, 50%, and 25% dilutions of Sal_Hartnel WW. The calculated LC50 was 26.29% (20.93, 32.04), with a NOEC of 12.5% and a LOEC of 25%. There were 3.8 Toxic Units in this sample. No significant reductions in survival were observed in any other treatment. Toxicity test results are presented below in Tables 1-4. Water quality measurements from these tests are outlined in Tables 5-8.

Table 1. Summary of results from a *H. azteca* toxicity test initiated on December 1, 2021 with whole water samples collected from Sal-Hartnel on November 29, 2021.

Sample	24-1	24-hr survival		48-ŀ	48-hr survival			72-hr Survival			96-hr Survival		
Sample	Mean	SD	SE										
0% (Control)	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	
6.25% Sal_Hartnel WW	100.0	0.00	0.00	98.0	4.47	2.00	98.0	4.47	2.00	96.0	5.48	2.45	
12.5% Sal_Hartnel WW	100.0	0.00	0.00	98.0	4.47	2.00	92.0	17.89	8.00	88.0	21.68	9.70	
25% Sal_Hartnel WW	98.0	4.47	2.00	90.0	12.25	5.48	82.0	13.04	5.83	62.0	8.37	3.74	
50% Sal_Hartnel WW	36.0	11.40	5.10	16.0	8.94	4.00	4.0	5.48	2.45	4.0	5.48	2.45	
100% Sal_Hartnel WW	26.0	15.17	6.78	14.0	8.94	4.00	2.0	4.47	2.00	2.0	4.47	2.00	

^{1.} Highlighted cells indicate a statistically significant reduction (*P*<0.05) in survival compared to the control.

Table 2. Summary of results from a *H. azteca* toxicity test initiated on December 1, 2021 with clarified supernatant samples collected from Sal-Hartnel on November 29, 2021.

Sample	24-h	24-hr survival		48-hr survival			72-hr Survival			96-hr Survival		
Sample	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE
0% (Control)	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00
6.25% Sal_Hartnel CS	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00
12.5% Sal_Hartnel CS	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00
25% Sal_Hartnel CS	100.0	0.00	0.00	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00
50% Sal_Hartnel CS	100.0	0.00	0.00	100.0	0.00	0.00	98.0	4.47	2.00	98.0	4.47	2.00
100% Sal_Hartnel CS	100.0	0.00	0.00	96.0	5.48	2.45	94.0	5.48	2.45	94.0	5.48	2.45

Table 3. Summary of results from a *H. azteca* toxicity test initiated on December 1, 2021 with whole water samples collected from Sal-Tembl on November 30, 2021.

Sample	24-1	24-hr survival		48-hr survival			72-hr Survival			96-hr Survival		
Sample	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE
0% (Control)	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00
6.25% Sal Tembl WW	100.0	0.00	0.00	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00

Sample	24-hr survival		48-ł	48-hr survival			72-hr Survival			96-hr Survival		
Sample	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE	Mean	SD	SE
12.5% Sal_Tembl WW	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	98.0	4.47	2.00
25% Sal_Tembl WW	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00
50% Sal_Tembl WW	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00
100% Sal_Tembl WW	100.0	0.00	0.00	100.0	0.00	0.00	94.0	13.42	6.00	94.0	13.42	6.00

Table 4. Summary of results from a *H. azteca* toxicity test initiated on December 1, 2021 with clarified supernatant samples collected from Sal-Tembl on November 30, 2021.

Sample	24-ŀ	24-hr survival		48-ŀ	48-hr survival			72-hr Survival			96-hr Survival		
Sample	Mean	SD	SE										
0% (Control)	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	
6.25% Sal_Tembl CS	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	98.0	4.47	2.00	
12.5% Sal_Tembl CS	100.0	0.00	0.00	100.0	0.00	0.00	96.0	5.48	2.45	98.0	4.47	2.00	
25% Sal_Tembl CS	98.0	4.47	2.00	98.0	4.47	2.00	98.0	4.47	2.00	100.0	0.00	0.00	
50% Sal_Tembl CS	100.0	0.00	0.00	96.0	5.48	2.45	96.0	5.48	2.45	100.0	0.00	0.00	
100% Sal_Tembl CS	100.0	0.00	0.00	100.0	0.00	0.00	100.0	0.00	0.00	94.0	13.42	6.00	

Table 5. Summary of water quality measurements from a *H. azteca* toxicity test initiated on December 1, 2021 with whole water samples collected from Sal_Hartnel on November 29, 2021.

Cample		DO (mg/L)		Initial Co	C/uS/cm)
Sample	Initial	Final	Min	Max	IIIIIIai SC	C (µS/cm)
0% (Control)	8.78	7.67	7.67	8.78	3	58
6.25% Sal_Hartnel WW	8.58	7.64	7.60	8.58	4	15
12.5% Sal_Hartnel WW	8.88	7.54	7.54	8.88	450	
25% Sal_Hartnel WW	8.78	7.55	7.55	8.78	5	34
50% Sal_Hartnel WW	8.98	7.23	7.23	8.98	6	99
100% Sal_Hartnel WW	8.25	7.21	7.21	8.25	10)34
		p	Н		Total	Unionized
Sample	Initial	Final	Min	Max	Ammonia (mg/L)	Ammonia (mg/L) ¹
0% (Control)	7.74	7.60	7.58	7.94	ND	ND
6.25% Sal_Hartnel WW	7.70	7.69	7.51	7.93	NR	NR
12.5% Sal_Hartnel WW	7.70	7.59	7.48	7.96	NR	NR
25% Sal_Hartnel WW	7.69	7.64	7.54	7.96	NR	NR
50% Sal_Hartnel WW	7.66	7.69	7.63	7.99	NR	NR
100% Sal_Hartnel WW	7.67	7.82	7.67	7.99	2.58	0.052
		Tem	p (°C)		Alkalinity	Hardness
Sample	Initial	Final	Min	Max	(mg/L as CaCO₃)	(mg/L as CaCO₃)
0% (Control)	23.4	22.2	22.2	24.3	54	100
6.25% Sal_Hartnel WW	23.5	22.0	22.0	24.3	NR	NR
12.5% Sal_Hartnel WW	23.4	22.2	22.2	24.5	NR	NR
25% Sal_Hartnel WW	23.4	22.1	22.1	24.3	NR	NR
50% Sal_Hartnel WW	23.3	22.2	22.2	24.3	NR	NR
100% Sal_Hartnel WW	23.4	22.1	21.4	24.3	150	376

^{1:} This unionized ammonia reading is based on the total ammonia measured upon sample receipt and upon the water chemistry measured at test initiation. NR: Ammonia, hardness, and alkalinity were measured on the control and the 100% concentration of the ambient sample only. ND: Non-detect.

Table 6. Summary of water quality measurements from a *H. azteca* toxicity test initiated on December 1, 2021 with clarified supernatant samples collected from Sal_Hartnel on November 29, 2021.

Sample		DO (ı	mg/L)		Initial SC	C (μS/cm)
Sample	Initial	Final	Min	Max	illitiai 30	ζ (μ3/τιτι)
0% (Control)	8.83	7.72	7.72	8.83	3	55
6.25% Sal_Hartnel CS	8.56	8.00	7.76	8.56	4	11
12.5% Sal_Hartnel CS	8.83	7.83	7.77	8.83	4	61
25% Sal_Hartnel CS	9.09	8.00	7.84	9.09	5	41
50% Sal_Hartnel CS	9.15	8.11	7.77	9.15	7	50
100% Sal_Hartnel CS	8.90	8.24	7.68	8.90	10)54
		р	Н		Total	Unionized
Sample	Initial	Final	Min	Max	Ammonia (mg/L)	Ammonia (mg/L) ¹
0% (Control)	7.72	7.58	7.58	7.87	ND	ND
6.25% Sal_Hartnel CS	7.70	7.64	7.64	7.89	NR	NR
12.5% Sal_Hartnel CS	7.70	7.63	7.63	7.92	NR	NR
25% Sal_Hartnel CS	7.70	7.79	7.70	7.94	NR	NR
50% Sal_Hartnel CS	7.70	7.88	7.70	8.04	NR	NR
100% Sal_Hartnel CS	7.68	8.02	7.63	8.08	0.17	0.004
		Tem	p (°C)		Alkalinity	Hardness
Sample	Initial	Final	Min	Max	(mg/L as CaCO₃)	(mg/L as CaCO₃)
0% (Control)	23.4	22.1	22.1	24.3	54	100
6.25% Sal_Hartnel CS	23.6	21.9	21.9	24.3	NR	NR
12.5% Sal_Hartnel CS	23.5	22.0	21.7	24.2	NR	NR
25% Sal_Hartnel CS	23.6	22.0	21.7	24.1	NR	NR
50% Sal_Hartnel CS	23.5	21.9	21.9	24.0	NR	NR
100% Sal_Hartnel CS	23.6	21.8	21.8	24.0	134	384

^{1:} This unionized ammonia reading is based on the total ammonia measured upon sample receipt and upon the water chemistry measured at test initiation. NR: Ammonia, hardness, and alkalinity were measured on the control and the 100% concentration of the ambient sample only. ND: Non-detect.

Table 7. Summary of water quality measurements from a *H. azteca* toxicity test initiated on December 1, 2021 with whole water samples collected from Sal_Tembl on November 30, 2021.

Sample		DO (ı	mg/L)		Initial SC	C (µS/cm)
Sample	Initial	Final	Min	Max	illitiai 30	ζ (μ3/τιτι)
0% (Control)	8.83	7.68	7.68	8.83	35	7.5
6.25% Sal_Tembl WW	9.03	7.77	7.67	9.03		
12.5% Sal_Tembl WW	9.19	7.78	7.74	9.19	587	
25% Sal_Tembl WW	9.20	8.22	7.69	9.20	7	86
50% Sal_Tembl WW	9.24	8.40	7.74	9.24	11	L97
100% Sal_Tembl WW	9.29	8.66	7.70	9.29	19	962
		р	Н		Total	Unionized
Sample	Initial	Final	Min	Max	Ammonia (mg/L)	Ammonia (mg/L) ¹
0% (Control)	7.69	7.59	7.56	7.94	ND	ND
6.25% Sal_Tembl WW	7.73	7.7	7.67	8.13	NR	NR
12.5% Sal_Tembl WW	7.79	7.81	7.72	8.05	NR	NR
25% Sal_Tembl WW	7.84	7.97	7.84	8.14	NR	NR
50% Sal_Tembl WW	7.87	8.23	7.87	8.23	NR	NR
100% Sal_Tembl WW	7.89	8.37	7.89	8.37	0.098	0.003
		Tem	p (°C)		Alkalinity	Hardness
Sample	Initial	Final	Min	Max	(mg/L as CaCO₃)	(mg/L as CaCO₃)
0% (Control)	23.6	22.1	22.1	24.1	54	100
6.25% Sal_Tembl WW	23.6	21.9	21.9	24.2	NR	NR
12.5% Sal_Tembl WW	23.6	22.1	22.1	24.1	NR	NR
25% Sal_Tembl WW	23.6	21.9	21.9	24.1	NR	NR
50% Sal_Tembl WW	23.6	22.2	22.2	24.0	NR	NR
100% Sal_Tembl WW	23.6	22.1	22.1	23.8	296	720

^{1:} This unionized ammonia reading is based on the total ammonia measured upon sample receipt and upon the water chemistry measured at test initiation. NR: Ammonia, hardness, and alkalinity were measured on the control and the 100% concentration of the ambient sample only. ND: Non-detect.

Table 8. Summary of water quality measurements from a *H. azteca* toxicity test initiated on December 1, 2021 with clarified supernatant samples collected from Sal_Tembl on November 30, 2021.

Sample		DO (ı	mg/L)		Initial SC	C (μS/cm)
Sample	Initial	Final	Min	Max	illitiai 30	υ (μο/ τιτι)
0% (Control)	8.89	7.66	7.66	8.89	3	54
6.25% Sal_Tembl CS	8.82	8.86	7.53	8.86	4	79
12.5% Sal_Tembl CS	9.00	7.78	7.60	9.00	5	74
25% Sal_Tembl CS	9.11	7.90	7.66	9.11	7	94
50% Sal_Tembl CS	8.95	7.75	7.68	8.95	11	189
100% Sal_Tembl CS	9.22	7.64	7.59	9.22	19	961
		р	Н		Total	Unionized
Sample	Initial	Final	Min	Max	Ammonia (mg/L)	Ammonia (mg/L) ¹
0% (Control)	7.67	7.64	7.59	7.92	ND	ND
6.25% Sal_Tembl CS	7.76	7.70	7.66	8.04	NR	NR
12.5% Sal_Tembl CS	7.80	7.79	7.75	8.06	NR	NR
25% Sal_Tembl CS	7.87	7.91	7.85	8.15	NR	NR
50% Sal_Tembl CS	7.90	8.13	7.90	8.22	NR	NR
100% Sal_Tembl CS	7.91	8.27	7.91	8.32	ND	ND
		Tem	p (°C)		Alkalinity	Hardness
Sample	Initial	Final	Min	Max	(mg/L as CaCO₃)	(mg/L as CaCO₃)
0% (Control)	23.7	22.1	22.1	24.1	54	100
6.25% Sal_Tembl CS	23.7	22.2	22.2	24.3	NR	NR
12.5% Sal_Tembl CS	23.6	22.3	22.3	24.4	NR	NR
25% Sal_Tembl CS	23.6	22.4	22.4	24.4	NR	NR
50% Sal_Tembl CS	23.5	22.3	22.3	24.4	NR	NR
100% Sal_Tembl CS	23.5	22.0	22.0	24.4	286	720

^{1:} This unionized ammonia reading is based on the total ammonia measured upon sample receipt and upon the water chemistry measured at test initiation. NR: Ammonia, hardness, and alkalinity were measured on the control and the 100% concentration of the ambient sample only. ND: Non-detect.

Table 9. Summary of Work Completed To Date.

Task	Items for Review #	% Complete	Estimated Due Date	Work Completed
	1: Coordination of sample collection	50	Ong	oing
Task 1	1.1 Collection of whole water	50	May 30, 2022	
	1.2 Collection of clarified supernatant	50	May 30, 2022	
	2: H. azteca toxicity testing: WW + CS	50	Ong	oing
	2.1 Central Coast Location 1	100	Nov. 2021	Dec. 1, 2021
Task 2	2.2 Central Coast Location 2	100	Nov. 2021	Dec. 1, 2021
	2.3 Central Coast Location 3	0	May 2022	
	2.3 Central Coast Location 4	0	May 2022	
	3: H. azteca settling column tests	0	Ong	oing
	3.1 Supernatant: Suspended solids	0	April 2022	
Task 3	3.2 Re-suspended supernatant: suspended solids	0	April 2022	
	3.3 Separated supernatant: suspended solids	0	April 2022	
	3.4 Post-test analytical chemistry	0	April 2022	
Task 4	4: Toxicity information to mathematical model	0	Ong	oing
Task 5	5: Application of model to field scale sites	0	Ong	oing
	6: Administration and Reporting	6	Ong	oing
	6.1 Quarterly Reports	6	Feb. 1, 2023	Feb. 15, 2022
Task 6	6.2 Quarterly Invoices	6	Jun. 30, 2023	Ongoing
	6.3 Draft Project Report	0	Apr. 15, 2023	
	6.6 Final Report	0	Jun. 30, 2023	

Literature Cited

- 1. Aquatic Health Program. 2020. Standard Operating Procedures. University of California, Davis. Davis, CA.
- 2. Surface Water Ambient Monitoring Program. 2008. Quality Assurance Project Plan. For the State Water Resources Control Board. Sacramento, CA.
- 3. USEPA, 2002. Short-Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms. 4th ed. *EPA/821/R-02/013*. Office of Water. Washington, DC.